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SYNTHESIS AND PHARMACOLOGICAL EVALUATION OF HIGHLY POTENT DUAL HISTAMINE H₂ AND GASTRIN RECEPTOR ANTAGONISTS

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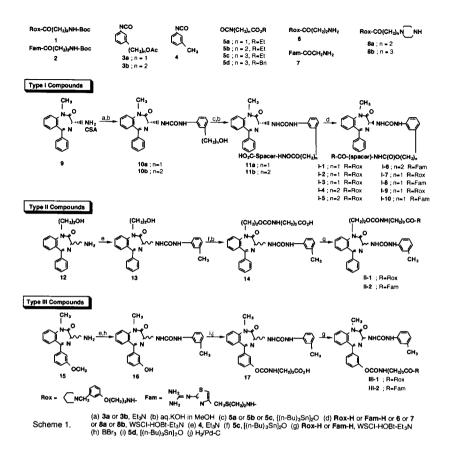
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Abstract: The chemical modification of the dual histamine H₂ and gastrin receptor antagonists described in our preceding paper, particularly the modification of spacers as well as the alteration of their connecting bonds at the gastrin receptor antagonist site (GA) from the amide bond to the carbamate bond, significantly improved not only their dual activity but also the GA versus CCK-A receptor selectivity. Copyright © 1996 Elsevier Science Ltd

With the objective of alleviating the relapse problem frequently encountered in the chemotherapy of peptic ulceration with histamine H₂ receptor antagonists (H₂A), we reported in our preceding paper ¹ the design, synthesis, and pharmacological evaluation of dual histamine H₂ and gastrin receptor antagonists. In that study, we designed as the dual antagonist, a joint type of hybrid molecule composed of a H₂A moiety and a GA one which were selected from well-known H₂As, famotidine and roxatidine, and GA L-365,260, respectively. We found that to retain the dual activity, it was important to consider the spacers as well as the three sites of L-365,260 to which the H₂A moieties could be attached, namely the N₁ methyl group, the *meta* methyl group on the C₃ tolyl group, and the *meta* position of the phenyl group at C₅. Consequently, we obtained several compounds with the dual activity. However, they showed distinct disadvantages such as lower H₂A and GA activities compared to famotidine and L-365,260, respectively, smaller gastrin versus CCK-A receptor selectivities than that of L-365,260, and decreased oral absorbability. To improve these compounds, we tried altering the connecting bond of the spacers at the GA sites from the amide bond to the carbamate bond as well as modifying the structure of the spacers.

We chose the carbamate bond as the binding function of the spacers at the GA sites in this study for three reasons. First, as we had observed the extremely high pA2 values, 7.8 and 7.3, of H2A intermediate compounds 1 and 2 bearing the carbamate group, we predicted that the carbamate group, if properly located, would greatly potentiate the H2A activity of the hybrid compounds as well. Second, as the carbamate bond is less liable to enzymatic hydrolysis than the amide bond, its use as a binding function was expected to enhance the metabolic stability of hybrid molecules. Third, as the carbamate group is more hydrophobic than the amide group, its use might enhance the oral absorbability of the hybrid compounds.

The carbamate-type hybrid compounds² synthesized in this work are summarized in Scheme 1. These compounds can be classified into three types (Type I, II, III) depending upon the H₂A connecting sites at L-365,260 as previously described. The general synthetic scheme is briefly summarized in Scheme 1. First, isocyanates such as 3a, b and 4 were prepared using known methods. For the synthesis of Type I compounds, 3a and 3b were coupled with 3R-(+)-amino-1,3-dihydro-1-methyl-5-phenyl-2H-1,4-benzodiazepine-2-one camphorsulfonic acid salt (CSA salt) 9³ followed by alkaline hydrolysis to afford



the alcohol derivatives, 10a and 10b. For the syntheses of Type II and Type III compounds, the corresponding benzodiazepine amines, 12 and 15, were prepared as racemates using essentially the same methods as those reported and condensed with isocyanate 4 to afford the corresponding alcohol derivatives, 13 and 16, respectively. All the alcohols 10a, b, 13, 16 thus far prepared were allowed to react in the presence of bis tri-n-butyltin oxide catalysis with various spacer-coupled isocyanates 5a - d which were easily prepared by known methods. The resultant products were hydrolyzed under alkaline conditions to afford intermediate carbamate derivatives 11a, b, 14, and 17. These intermediates were subjected to coupling reactions with H2A amine moieties, Rox-H, Fam-H, 6, 7, 8a, or 8b, in the same way as previously reported, producing the desired spacers containing carbamate-type hybrid molecules as described above. All the hybrid compounds prepared with simple alkylene spacers are presented in Table 1 together with their in vitro biological activities. The significant features of their biological results are as follows: 1) Alteration of the spacer connecting bond from the amide bond to the carbamate bond significantly improved both the H2A and the GA activity of Type I hybrid compounds, particularly the latter. However, Type II and Type III compounds showed no or if any, only marginal, augmentation of these activities. 2) Most notably, the gastrin versus CCK-A receptor selectivity was highly improved by this structural modification (see Entry 3 vs 15). 3) The roxatidine type of hybrid compounds tended to show a higher pA2 value than their famotidine counterparts.

In order to examine pharmacological effects of spacers with polar functional groups such as amino acid glycine or the piperazine group, hybrid compounds I-7, I-8, I-9, I-10 were prepared. Their structures and biological activities are summarized in Table 1. The characteristics of the biological data are as follows: 1) Among these hybrid compounds, compound I-7 with a glycine type of spacer showed the highest pA2 value of 6.8 which obviously exceeded the value of cimetidine, 6.6. Its GA activity as well as gastrin versus CCK-A receptor selectivity were much better than those of the propylene compounds I-3 and I-5. Although its pA2 value is obviously lower than I-7, the famotidine type analogue I-8 shows the highest GA activity, IC50 = 4 nM, which is comparable to that of L-365,260. 2) The compounds with piperadine type spacers I-9 and I-10 show somewhat lower pA2 values, 6.2 and 6.5, than compounds I-7 and I-8 but their GA activities are retained at quite high levels, being IC50 = 7 nM and 9 nM, respectively. Thus, the most characteristic feature of the biological results here was the enhancement of both *in vitro* H2A and GA activities as well as the gastrin versus CCK-A receptor selectivity, the latter being approximately 100-fold better than that of the amide-type hybrid compounds.

Table 1.	In	Vitro	Biological	Activities
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Fater	No.	(CH ₂) _n	GA ← spacers → H ₂ B	H ₂ B (R,S)		Receptor	s IC ₅₀	(nM) G	Ratio	pA ₂
Entry		n≖				Gastrin	сск-в	CCK-A	CCK-A	His. H ₂
1	I-1	1	-CH ₂ -	Rox	R	11	200	6100	555	6.1
2	l-2	1	-CH ₂ CH ₂ -	Rox	R	36	160	310	9	6.5
3	I-3	1	-CH ₂ CH ₂ CH ₂ -	Rox	R	26	94	3600	138	6.6
4	I-4	2	-CH ₂ CH ₂ -	Rox	R	27	94	4200	156	5.9
5	I-5	2	-CH ₂ CH ₂ CH ₂ -	Rox	R	23	180	5000	217	6.4
6	I-6	2	-CH ₂ CH ₂ CH ₂ -	Fam	R	37	300	1300	35	6.1
7	II-1		-CH ₂ CH ₂ CH ₂ -	Rox	RS	135	4000	4600	34	6.0
8	II-2		-CH ₂ CH ₂ CH ₂ -	Fam	RS	17	930	4400	289	5.5
9	III-1		-CH ₂ CH ₂ CH ₂ -	Rox	RS	220	460	3800	17	6.3
10	III-2		-CH ₂ CH ₂ CH ₂ -	Fam	RS	>1000	>1000	>10000	10	5.7
11	ŀ 7	1	-CH ₂ CONHCH ₂ CH ₂ -	Rox	R	19	103	8200	432	6.8
12	I-8	1	-CH ₂ CONHCH ₂ -	Fam	R	4	75	3100	775	6.0
13	I-9	1	-H2COCN_NCH2CH2-	Rox	R	7	45	740	106	6.2
14	l-10	1	-H2COCN_NCH2CH2CH2	- Rox	R	9	400	1650	183	6.5
15	Amide type	1	-CH ₂ CH ₂ CH ₂ -	Rox	R	115	205	330	3	6.4
16	1			Rox						7.8
17	2			Fam						7.3
18 19	Cimetic L-365,2				R	4	29	11100	2775	6.6

The *in vivo* gastric acid antisecretory activity of the most promising compound **I-7** was evaluated by three different methods, namely Schild's rat method⁴ (i.d. route administration), rat pylorus ligation method⁵ (p.o.), and canine Heidenhain pouch method⁶ (i.v. and p.o.). When **I-7** was administered to Heidenhain pouch dog by i.v. route, it distinctly inhibited gastric acid secretion by 51% at 0.3 mg/kg dose. Nevertheless, it did not show any notable inhibition even at 30 mg/kg dose via oral administration. For Schild's rat method and rat pylorus ligation method, it inhibited histamine-stimulated gastric acid secretion only 77% at 30 mg/kg dose and less than 50% at 10 mg/kg dose in the former and only 54% at 10 mg/kg

dose in the latter. All these results suggested low oral absorbability of this compound in these animal species. Next, the *in vivo* GA activity of I-7 was evaluated as the % inhibition of tetragastrin-induced histidine decarboxylase activity. Inhibition was statistically positive but low, being 32% at 30 mg/Kg dose by i.p. route. Because of the low oral absorbability of I-7, we could not evaluate its pharmacological effects as a typical compound for suppressing the rebound of gastric acid after cessation and thereby relapse of ulcer. Although the species dependency of the GA activity as well as the diminished oral absorbability of L-365,260 in higher animals have been documented 7, the present study strongly indicated that improvement of their diminished oral absorbability would be essential for the development of these novel dual histamine H₂ and gastrin antagonists as antiulcer agents for practical use.

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- 2. For a typical example of the general experimental procedure for the synthesis of hybrid molecules refer to the preceding paper. All three types of hybrid compounds were obtained in 50 - 70% yield by this method and gave satisfactory analytical data on elemental analysis. ¹H NMR data [200 MHz. 8 (ppm), J (Hz)] for selected hybrid compounds with glycine or piperadine spacers: I-5; (CD3OD) 1.60-2.05 (m, 8H, 4CH₂), 2.16 (t, 2H, CH₂, J = 6 Hz), 2.80-3.00 (m, 4H, 2CH₂), 3.08 (t, 2H, CH₂, J = 6 Hz), 2.80-3.00 (m, 4H, 2CH₂), 3.08 (t, 2H, CH₂, J = 6 Hz), 2.80-3.00 (m, 4H, 2CH₂), 3.08 (t, 2H, CH₂, J = 6 Hz), 2.80-3.00 (m, 4H, 2CH₂), 3.08 (t, 2H, CH₂, J = 6 Hz), 2.80-3.00 (m, 4H, 2CH₂), 3.08 (t, 2H, CH₂, J = 6 Hz), 3.08 (t, 2H, CH₂), 3.086 Hz), 3.30-3.60 (m, 6H, 3CH₂), 3.51 (s, 3H, N-CH₃), 4.03 (t, 2H, CH₂, J = 6 Hz), 4.17 (s, 2H, $\overline{CH_2}$), 4.17 (t, 2H, CH₂, J = 6 Hz), 5.37 (s, 1H, 3-H), 6.85-7.80 (m, 17H, Ar-H). I-6; 1.60-1.80 (m, 2H, CH₂), 2.07-2.22 (m, 2H, CH₂), 2.57 (t, 2H, CH₂, J = 6 Hz), 2.85 (t, 2H, CH₂, J = 6 Hz), 3.07 (t, 2H, CH₂) CH₂, J = 6 Hz), 3.25-3.37 (m, 2H, CH₂), 3.49 (s, 3H, CH₃), 3.63 (s, 2H, CH₂), 4.20 (t, 2H, CH₂, J = 6 Hz), 5.37 (s, 1H, 3-H), 6.52 (s, 1H, thiazole-CH), 6.85-7.75 (m, 13H, Ar-H). I-7; (CDCl₃) 1.35-1.66 (m, 6H, 3CH₂), 1.79-1.96 (m, 2H, CH₂), 2.36-2.56 (m, 6H, 3CH₂), 3.31-3.42 (m, 2H, CH₂), 3.40 (s, 3H, CH₃), 3.50 (s, 2H, CH₂), 3.75 (d, 2H, CH₂), 3.88-3.96 (m, 2H, CH₂), 5.00 (s, 2H, CH₂), 3.50 (d, 1H, 3-H, J = 8 Hz), 6.69-7.60 (m, 17H, Ar-H). I-9; (CD3OD) 1.35-1.68 (m, 6H, 3CH2), 1.69-1.88 (m, 2H, CH₂), 1.88-2.04 (m, 2H, CH₂), 2.24 (m, 2H, CH₂), 2.22-2.38 (m, 8H, 4CH₂), 2.37-2.58 (m, 4H, 2CH₂), 3.18-3.36 (m, 2H, CH₂), 3.45 (s, 3H, CH₃), 3.35-3.52 (m, 2H, CH₂), 3.55 (s, 2H, CH₂), 3.95 (d, 2H, CH₂, J = 5.6 Hz), 4.05 (m, 2H, CH₂), 5.04 (s, 2H, CH₂), 5.54 (d, 1H, 3-H, J = 8.2 Hz), 6.74-7.62 (m, 13H, Ar-H). II-1; (CD₃OD) 1.40-1.70 (m, 8H), 1.98 (m, 4H), 2.28 (s, 3H), 2.40 (m, 4H), 2.83 (m, 2H), 3.34 (t, 2H, J = 7 Hz), 3.45 (s, 2H), 3.98 (t, 2H, J = 7 Hz), 3.90-4.20 (m, 3H), 4.71 (m, 1H), 5.38 (s, 1H), 6.75-7.80 (m, 17H). III-1; (CD₃OD) 1.35-1.68 (m, 6H, 3CH₂), 1.69-1.88 (m, 2H, CH₂), 1.88-2.04 (m, 2H, CH₂), 2.24 (m, 2H, CH₂), 2.22-2.38 (m, 8H, 4CH₂), 2.37-2.58 (m, 4H, 2CH₂), 3.18-3.36 (m, 2H, CH₂), 3.45 (s, 3H, CH₃), 3.35-3.52 (m, 2H, CH₂), 3.55 (s, 2H, CH₂), 3.95 (d, 2H, CH₂, J = 5.6 Hz), 4.05 (m, 2H, CH₂), 5.04 (s, 2H, CH₂), 5.54 (d, 1H, 3-H, J = 8.2 Hz), 6.74-7.62 (m, 13H, Ar-H).
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